Modeling Tumor Regrowth and Immunotherapy

Vladimir A. Kuznetsov, Gary D. Knott

Civilized Software, Inc. 12109 Heritage Park Circle Silver Spring MD 20906 Tel.: (301)-962-3711 email: csi@civilized.com URL: www.civilized.com

Introduction

Tumors that originate in the body via a variety of mechanisms usually grow slowly for some time. Many months or years may be needed for the existence of such a tumor to manifest itself. This near-steady-state existence of a tumor is described by the term *cancer dormancy* [Uhr et al., 1997][Vitetta et al., 1997]. Cancer dormancy is a well-recognized clinical phenomenon in which malignant cells persist for a prolonged period of time with little or no increase in the tumor cell population. This state may occur naturally, or following apparently-effective therapy.

There are at least two plausible independent pathways to the clinically "quiescent state" of a tumor. The first pathway corresponds to intrinsic properties of the tumor cells (related to the expression of suppressor genes, production of growth and/or anti-growth factors and corresponding receptors, etc.) The second pathway corresponds to approaching an equilibrium of interaction between the growing tumor cell population and various cellular and molecular components of the immune system. In either or both situations, the tumor appears to be dormant. Nevertheless, tumor dormancy is not necessarily a stable state. Many factors - infection, stress, immunodepression events, changes in endocrine status, etc. may disturb the balance between the host and the dormant tumor.

We will present a mathematical model for tumor growth and suppression below and show that this model can describe the regrowth of a dormant tumor by either of two distinct mechanisms. The first mechanism is a modest decrease in the efficiency of immune-suppression of the tumor which has, prior to this decrease, been able to maintain an equilibrium state wherein the tumor does not exhibit growth. Variants of this mechanism have been studied in [Kuznetsov, 1991] and [Kuznetsov et al., 1994]. The second mechanism we explore is the existence of a mutant clone of tumor cells which is not effectively suppressed by an immune response and hence grows unchecked, except for natural constraints imposed by nutrient resources.

In our model, tumor growth is reversed and then held in check by the continual attack of killer cells; however, when the equilibrium relationship between the killer cell and the tumor cell populations is suitably perturbed, the tumor "escapes" control and grows.

Single Clone Model for Tumor Regrowth

Let c(t) denote the number of cancer cells present at time t, measured in units of 1 million cells. Let e(t) denote the number of cytotoxic killer cells present at time t, measured in units of 1 million cells. Then the growth-rate of the cancer cell population is defined by the differential equation:

$$c'(t) = kc(t)(1 - bc(t)) - ape(t)c(t).$$

The term kc(1-bc) is the intrinsic rate of tumor cell growth. The parameter k is the maximal rate of tumor-cell growth (when b = 0, the tumor grows at the rate k,) and the parameter b is the reciprocal of the maximal tumor cell population size; *i.e.* 1/b = (the maximum number of tumor cells permitted to arise) $/10^{6}$.

The term *apec* specifies the rate of destruction of tumor cells. We assume that killer cells bind with tumor cells, and that when this occurs then either (1) the tumor cell is destroyed, or (2) the killer cell is destroyed or inactivated. (The event that both cells are destroyed is deemed negligible.)

The parameter p is the probability that a tumor cell bound with a killer cells will be destroyed, and 1 - p is the probability that a tumor-cell-killer-cell bound pair results in the killer cell being destroyed and the tumor cell surviving. The parameter a is defined so that a(1-p) is the rate of killer cell destruction (after binding with a tumor cell,) and ap is the rate of tumor cell destruction (after binding with a killer-cell.) Note that 1 - a is the rate of neither cell killing the other after binding, and unbinding to try again. Thus, ap is the "kinetic" constant that multiplies e and c to form the overall tumor-cell destruction rate term apec.

The growth-rate of the killer-cell population (which changes size as new killer cells are attracted and arrive via the lymph system) is defined by the differential equation:

$$e'(t) = r + (f(t)c(t)/(g+c(t)))e(t) - de(t) - (a(1-p)c(t))e(t)$$

The parameter r is the base rate of arrival of killer cells at the tumor via the lymph system; we take r to be a derived parameter defined as e_0d , where $e_0 = .3$ (.3 million is our estimate of the number of killer cells present in the absence of tumor cells taken from [Kuznetsov et al., 1994]. The term (f(t)c/(g+c))e is the additional rate of both local proliferation and arrival of killer cells due to the chemokine and cytokine stimulation signals induced by the tumor-cell population of size c.

The expression f(t)c/(g+c) is a logistic growth rate expression in which f(t) = if t = 0 then v else if t < u then 0 else v. (The notation "if D then E_1 else E_2 " defines a function with a discontinuity, so that (if D then E_1 else E_2) is the value of the expression E_1 when the condition D holds, and is the value of the expression E_2 otherwise.) The parameter v is the maximal rate of logistic growth of killer-cell population due to tumor growth, and the parameter u is the delay time of the immune response before "new" killer cells can be applied to attack the tumor, *i.e.* u is the time for precursor cells to mature into killer cells. The function f "cancels" its additional growth term until t > u. The parameter g is the mid-point (IC-50) logistic parameter in the additional arrival rate term. The parameter d is the natural rate of death of killer cells. Finally, the term a(1-p)ce is the rate of killer-cell death or inactivation due to the presence of H_2O_2 , gangliosides, cytolytic peptides produced by the tumor, etc.

We have used the experimental data due to Sui et al. [Sui 1986], where BCL_1 lymphoma tumor cells were injected into the spleens of chimeric mice, and the resulting tumor growth was followed. In particular we are given the mean number of tumor cells seen at various times in four groups of mice (numbered 0,1,2,3). These seeded tumors were initially of approximate size .5 million cells per mouse in group 0, .5 million cells per mouse in group 1, 5 million cells per mouse in group 2, and 50 million cells per mouse in group 3. In groups 1, 2 and 3 the resulting tumors respond to the action of the immune system. Group 0 mice have no immune response; tumor-size data from this group will be used to help estimate the parameters that define tumor growth by fitting the pure growth model c'(t) = kc(t)(1 - bc(t)) with c(0) = .5.

It is appropriate to both fit and view the data and our fitted models in logarithmic form; doing this is equivalent to weighting the data to assume the error in the tumor-size observations is more nearly log-normal than normal. The main purpose, however, is to introduce convenient units for both fitting and viewing. In order to fit the data, we replicated our model for each of the group 1,2, and 3 data-sets, and introduced the pure-growth differential equation model mentioned just above for the group 0 data. We have assumed that the initial number of killer cells contained in the spleen of the chimeric mice of groups 1,2, and 3 is .3 million, so, we used the initial conditions $e(0) = e_0$ for each replicated differential equation, where $e_0 = .3$. This indicates that $e_0 10^6$ is the number of killer cells that were initially present when the tumor cells were injected.

We used the MLAB mathematical and statistical modeling system from Civilized Software Inc., Silver Spring, MD, (see *www.civilized.com*) to fit our models and draw the corresponding graphs, since MLAB is especially designed to deal with differential equation models, including large systems of stiff equations. (The differential equations being studied here are stiff.) For the four data-sets used, fitting our model consists of fitting four functions defined by seven differential equations to estimate the eight parameters d, u, v, p, a, k, b, and q. This fitting required various exploratory computations and careful search of the parameter space for suitable initial guesses that lead to physiologically-plausible values. Our result is d = .5910007682, u = 28.05445851, v = .524999404, p = .9982002827, a = .138698686, k =.1877015458, b = .001880059483, and g = .1607110637. Note particularly that 28 days is an appropriate estimate of the maturation time for $CD8^+$ cytotoxic T lymphocytes. Figure 1 below shows this fit. Note for groups 1, 2, and 3, the tumor growth is suppressed and enters a equilibrium state of apparent dormancy. The limiting stable steady state of our model in this situation may be interpreted as the tumor dormant state.

The MLAB do-file (script file) that produced the above figure is given below.

reset echodo=3

/* We are given the mean number of tumor cells in four groups (0,1,2,3) of mice at various times. These tumors were initially of size .5, .5, 5, 50 million cells per mouse in each group. In groups 1, 2 and 3 these tumors respond to the action of the immune system. Group 0 mice have no immune response; this group will be used to estimate the parameters that

```
define tumor growth.
   Our model consists of two functions:
    e(t)=(# cytotoxic killer cells at time t)/10<sup>6</sup>, and
    c(t)=(\# \text{ cancer cells at time } t)/10^{6}
   for groups 1,2, and 3, and just c(t) for group 0.
*/
/* no immune response group 0, initial size =.5,
   col 1 in units of days, col 2 in units of 10<sup>6</sup> cells. */
cOdata col 1 =list(0, 20, 30, 50, 60, 70, 95)
cOdata col 2 =list(0.5, 16, 135, 405, 440, 468, 513)
/* immune response group 1, initial size =.5 */
cldata col 1=list(0, 20, 35, 40, 50, 70, 90, 110)
c1data col 2=list(.5, 6.31, 63.1, 35.48, 28.18, 2.82, 0.89, 0.89)
/* immune response group 2, initial size =5 */
c2data col 1=list(0, 20, 30, 70)
c2data col 2=list(5, 100, 177.8, 11.22)
/* immune response group 3, initial size =50 */
c3data col 1 =list(0, 10, 20, 40, 60, 70, 90, 110)
c3data col 2 =list(50, 251.2, 398.2, 281.84, 63.1, 50, 3.16, 1.26)
/* transform data by taking log10 of # of cells */
fct l(x) = 6 + \log 10(x)
lcOd=(cOdata col 1)&'(l on (cOdata col 2))
lc1d=(c1data col 1)&'(l on (c1data col 2))
lc2d=(c2data col 1)&'(l on (c2data col 2))
lc3d=(c3data col 1)&'(l on (c3data col 2))
function 110(x)=6+log10(x+.000001)
function e1't(t)=.3*d +(f(t)*c1/(g+c1))*e1 -d*e1 -a*(1-p)*e1*c1
function c1't(t)=k*c1*(1-b*c1) -a*p*e1*c1
function lc1(t) = l10(c1)
function le1(t) = 110(e1)
function e2't(t)=.3*d +(f(t)*c2/(g+c2))*e2 -d*e2 -a*(1-p)*e2*c2
```

```
function c2't(t)=k*c2*(1-b*c2) -a*p*e2*c2
function lc2(t) = 110(c2)
function le2(t) = 110(e2)
function e3't(t)=.3*d +(f(t)*c3/(g+c3))*e3 -d*e3 -a*(1-p)*e3*c3
function c3't(t)=k*c3*(1-b*c3) -a*p*e3*c3
function lc3(t) = 110(c3)
function le3(t) = 110(e3)
fct f(t)=if t<u then 0 else v
function c0't(t)=k*c0*(1-b*c0) /* pure growth model */
function lcO(t) = 110(cO)
initial e1(0)=.3
initial c1(0)=.5
initial e2(0)=.3
initial c2(0)=5.
initial e3(0)=.3
initial c3(0)=50.
initial cO(0)=.5
/*parameter guesses (best-fit guesses used.) */
u= 28.05445851
d=.5910007682
v= .524999404
p=.9982002827
g=.1607110637
a=.138698686
k=.1877015458
b=.001880059483
errfac=.000001
method=gear
disastersw=-2
fit(k,b,u,d,v,p,g,a), lc0 to lc0d, lc1 to lc1d, lc2 to lc2d, lc3 to lc3d
```

```
/* draw the graphs */
m=points(lc0,le1,lc1,le2,lc2,le3,lc3, 0:200!166)
/* group 1 data & fit */
draw lc1d pt circle lt none color red
draw m col (1,3) lt dashed color red /* graph le1 vs. t*/
draw m col (1,4) color red /* graph lc1 vs. t*/
left title "log10(number of cells)"
bottom title "time in days"
top title "growth of tumor (initial size .5)"
frame .5 to 1, .5 to 1
w1=w
/* group 2 data and fit */
draw lc2d pt circle lt none color yellow
draw m col (1,5) lt dashed color yellow /* graph le2 vs. t*/
draw m col (1,6) color yellow /* graph lc2 vs. t*/
left title "log10(number of cells)"
bottom title "time in days"
top title "growth of tumor (initial size 5)"
frame .0 to .5, 0 to .5
w2=w
/* group 3 data and fit */
draw lc3d pt circle lt none color green
draw m col (1,7) lt dashed color green /* graph le3 vs. t*/
draw m col (1,8) color green /* graph lc3 vs. t*/
left title "log10(number of cells)"
bottom title "time in days"
top title "growth of tumor (initial size 50)"
frame .5 to 1, .0 to .5
w3=w
/* group 0 data & fit */
draw lcOd pt circle lt none color green
draw m col (1,2) color orange /* graph lc0 vs. t*/
left title "log10(number of cells)"
bottom title "time in days"
top title "tumor growth (initial size .5), no immune response"
```

```
frame .0 to .5, .5 to 1
w4=w
view
/* end of fig1.do */
```

Now, if we modify our model to have the parameter p change from 0.9982002827 to 90 percent of that value after 150 days, then tumor regrowth is exhibited as shown in Figure 2. This is done by replacing p in our seven differential equations by the expression h(t), and defining h(t) = if t < 150 then p else .9p. This result indicates that a modest decrease in the immune response effectiveness, corresponding to a small increase in the proportion 1 - p of killer lymphocytes being inactivated by tumor cells, dramatically changes the outcome of the disease.

It is worthwhile to note that suitably changing the immune response parameters v, d, or k can produce a similar regrowth event. However, increasing the parameter g from 0.16 to 5 did not change approaching the tumor dormancy steady state. Thus our modeling suggests that regrowth of dormant tumor may be associated with diminishing immune system activity, caused by a variety of mechanisms. Many immune functions are reduced with age or chronic stress. We explored the effect of diminishing the various immunological parameters in our model; we observed that slowly reducing the parameters p, v, d, or k with age also induced regrowth of the dormant tumor.

Two Clone Model for Tumor Regrowth

Another explanation of tumor regrowth is that there is a small population of immune-resistant cancer cells that are either initially-present or that develop and which grow unchecked by the action of killer cells. We can explore the behavior of such an aberrant clone population by introducing a pure growth model for such a population via the differential equation m'(t) = .33km(t)(1 - b(m(t) + c(t))) with m(0) = sc(0) and modifying the growth term kc(t)(1 - bc(t)) to be kc(t)(1 - b(m(t) + c(t))) in the differential equation that defines the function c. The parameter s is the proportion of the initial population of cells that matches the size of the initial population of aberrant immune-resistant cells.

Thus, the two clones model for cancer regrowth is written:

$$\begin{aligned} c'(t) &= kc(t)(1 - b(c(t) + m(t))) - ape(t)c(t) \\ m'(t) &= .33km(t)(1 - b(m(t) + c(t))), \\ e'(t) &= r + (f(t)c(t)/(g + c(t)))e(t) - de(t) - (a(1 - p)c(t))e(t), \end{aligned}$$

with $c(0) = c_0$, $m(0) = sc_0$, $e(0) = e_0$, and f(t) = if t < u then 0 else v, where $e_0 = .3$ and c_0 is variously equal to .5, 5, and 50.

We can estimate the value of s that causes the total tumor size c(t) + m(t) to rise to the value 500 after 290 days by simultaneously curve-fitting replicates of the function c(t) + m(t) to the point (290, 500) for groups 1,2, and 3, along with the data for each group of chimeric mice. We also fit these model functions to the additional data point (110, 1), corresponding to 10^6 being the number of dormant tumor cells present initially at days 100 to 110 as estimated in [Uhr et al., 1997]. MLAB permits such simultaneous fitting of many functions indirectly defined by differential equations. We obtained $s = 4.081604 \cdot 10^{-5} \pm 1.86 \cdot 10^{-5}$. The result is shown in Figure 3.

Simulation of Immunotherapy for an Established Dormant Cancer

Inducing an increased presence of various cytokines, chemokines and/or other immunomodulators in tumor tissue may augment the function of the immune system, and this can accomplished via vaccine agents.

In Figure 4 below, we show the results of numerically simulating the effects of changing the dynamics of the immune system via various vaccine regimes, thus modifying its action on an established dormant tumor. Such a change will result in reducing the tumor mass. However, generally this

response is temporary, and tumor regrowth may occur after stopping the immunotherapy and thereby reducing the influx of killer cells.

Figure 4a shows the result of increasing the flow of killer cells into a dormant tumor in three steps due to three imagined vaccine immunization treatments administered on days 180, day 194, and day 215. The modeled tumor is characterized by the parameters of a "normal" mouse obtained by fitting and exhibited in Figure 1. In this case, we assume the vaccination provokes a temporary rise in the size of the population of killer cells available to attack the tumor; this increase is independent of the (dormant) tumor-size. This is done by modifying the "base arrival rate" .3d by multiplying by factors that are temporarily greater than one, corresponding to each vaccine injection.

The therapeutic effect of vaccination shown in Figure 4a was modeled by the following equations.

$$\begin{aligned} e'(t) &= q(t)(.3d) + (f(t)c(t)/(g+c(t)))e(t) - de(t) - a(1-p)e(t)c(t), \text{ and} \\ c'(t) &= kc(t)(1-bc(t)) - ape(t)c(t), \text{ with} \\ f(t) &= \text{ if } t < u \text{ then } 0 \text{ else } v. \end{aligned}$$

The effects of the vaccinations are described by the "multiplier' function q, where:

$$\begin{aligned} q(t) &= f_1(t)f_2(t)f_3(t), \text{ and} \\ f_1(t) &= 1 + w(t, u_1), \\ f_2(t) &= 1 + w(t, u_2), \\ f_3(t) &= 1 + w(t, u_3), \text{ and} \\ w(t, z) &= \text{if } t < z \text{ then } 0 \text{ else } 60(\exp(-.35(t-z)) - \exp(-.4(t-z)). \end{aligned}$$

The function w(t, z) is 0 until time z; at time z it rises to a maximum of nearly 3 at about time z + 7 (indicating a 4-fold increase of the arrival-rate of killer cells); and thereafter exponentially decays to nearly 0 by day z + 18.

We used the initial conditions e(0) = .3, c(0) = 50 and the parameter values used for Figure 1, together with $u_1 = 180$, $u_2 = 194$, and $u_3 = 215$.

Figure 4b shows the result of modeling such imagined immunizations beginning ten days after tumor regrowth has started in a mouse with a diminished immune response as modeled in Figure 2; thus, the vaccinations occur at days 160, 174, and 195. Also, in this case, we assume our vaccine works by increasing the proliferation of killer cells in response to the logistic tumor size function, rather by increasing the arrival rate independently of the tumor size. This is done by multiplying the tumor-size dependent growth term for the killer cell population by factors which are temporarily greater than one, rather than modifying the base arrival rate as done above. Also, we assume that, although the number of killer cells increases in response to vaccination, their effectiveness, as measured by the parameter p, remains diminished at a level 10% lower than the "normal" mice modeled in Figure 1.

Our model is given by the following differential equations.

The effects of the vaccinations are here described by the "multiplier" function, q, defined in terms of the functions f_1 , f_2 , f_3 and w given above; f_1 , f_2 , f_3 , and w are defined as they were above, except that $u_1 = 160$, $u_2 = 174$, and $u_3 = 195$.

We used the initial conditions e(0) = .3, c(0) = 50 and the parameter values used for Figure 2, together with $u_1 = 160$, $u_2 = 174$, and $u_3 = 195$.

Figure 4c shows the result of our simulation of the effect of immunotherapy by injecting immune memory cells in a mouse with a regrowing tumor due to a diminished immune response as modeled in Figure 2 (and Figure 5b.) We assumed that, soon after injection, the memory cells will increase the immune response to the tumor due to enhanced stimulation of the production of killer cells for a period of about 150 days. This mechanism of adaptive immunotherapy is modeled with the differential equations:

$$\begin{aligned} e'(t) &= .3d + q(t)(f(t)c(t)/(g+c(t)))e(t) - de(t) - a(1-p)e(t)c(t), \text{ and} \\ c'(t) &= h(t)kc(t)(1-bc(t)) - ape(t)c(t), \text{ with} \end{aligned}$$

f(t) = if t < u then 0 else v, and

h(t) = if t < 150 then p else .9p.

The effects of the vaccinations are here again described by the "multiplier" function, q, defined in terms of the functions f_1 , f_2 , f_3 and w; f_1 , f_2 , and f_3 are defined as they were above, except that $u_1 = 160$, $u_2 = 174$, and $u_3 = 195$. The function w has the same form as before, except that it has been "lengthened" (by changing .35 to .35/10 and .4 to .4/10) to correspond

to an enhanced rate of proliferation of killer cells proportional to the logistic tumor size for a time period of about 150 days.

We used the initial conditions e(0) = .3, c(0) = 50 and the parameter values used for Figure 2, together with $u_1 = 160$, $u_2 = 174$, and $u_3 = 195$.

Figure 4d depicts the simulated situation where we modeled administering a "cytostatic" drug that has the effect of reducing the logistic growthrate parameter k that governs the rate of tumor regrowth; this might be an agent that reduces the tumors' nutrient supply, for example. In particular, we again used a reduced immune response mouse as described in Figure 2 which exhibits tumor regrowth starting at day 150. We imagine administering a drug that cuts the value of k in half on day 170. This is done mathematically by replacing k with the function $\lambda(t) = \text{if } t < 170$ then k else k/2.

Discussion

The assumption that, after the introduction of tumor cells (at time 0), no enhanced immune response occurred during the first 28 days after the mice were challenged with such injections was key to obtaining the excellent fits exhibited in Figure 1. The probability of inactivation of an immune killer cell after binding to a tumor cell is also a crucial parameter of the model. The value of p, as well as the numerical values of the other kinetic parameters of our model, are typical of the kinetic characteristics of CD8⁺ cytotoxic T lymphocytes involved in the allogenic immune response in mice.

We have studied two related mechanisms of tumor regrowth. The first model predicted that a small permanent reduction in the level of antitumor immune response may provoke the regrowth of a monoclonal tumor from a dormant state. Reducing the probability of killing or inactivation of a tumor cell by an immune killer lymphocyte, or reducing the rate of arrival of the immune lymphocytes into the tumor region, were the most critical factors in inducing the model to exhibit tumor regrowth. Various factors (*i.e.* aging, stress, infection, etc.) may explain why such a tumor-growthinducing change might occur.

The second two-clone tumor model also agreed with the experimental observations. Under the assumption that a small fraction (0.33%) of the tumor cell population injected initially into the mice is non-immunogenic and that the growth of this clone is three times slower than the growth rate of the major immunogenic tumor cell population, this model unsurprisingly predicted eventual tumor regrowth. Unlike our first model, This second model also predicts that size of the tumor after dormancy is determined by the initial number of injected tumor cells. Variability of initial doses of injected

cells explains the high dispersion of the time of clinical detection of regrown tumor after dormancy [Vitetta et al., 1997], under the assumptions of our second model. Moreover, immunological abnormality of a minor fraction of BCL_1 lymphoma cells was reported in the same study[Vitetta et al., 1997].

Note that the curves describing tumor regrowth presented above have distinct shapes. It would be interesting to see if tumor regrowth known to be due to immune system decline matches the regrowth profile in our first model while tumor regrowth due to the presence of a abnormal clone matches the regrowth profile of our second model.

Clinical and experimental observation confirms that intensive limitedterm immunotherapy does not provide complete tumor elimination, as predicted via modeling. Immunotherapy may reduce tumor mass to a handful of cells; however, if the functional activity of the immune system is slightly impared, tumor regrowth after immunization is likely. Model-fitting predicts that the life-time of killer cells is short (about 2 days). Medium-term control of cancer is exhibited when long-life immune memory cells are activated due to our simulated immunization. But long-term control results from reducing the cancer growth-rate.

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Figure 1: Fit of immune-response model to group 0-3 data. dashed = killer cells, solid = tumor cells, circles = data of Sui et al.



Figure 2: Fit of immune-response model to group 0-3 data with 10% lower killing probability after 150 days. dashed = killer cells, solid = tumor cells.



Figure 3: Anti-tumor immune response model with resistant aberrant clone fit to group 0-3 data with 10^6 tumor cells at day 110 and $500 \cdot 10^6$ tumor cells at day 290 for groups 1-3. dashed = killer cells, solid = tumor cells, long dashed = aberrant clone tumor cells.



Figure 4: Simulation of adjuvant immunization. (a) during dormancy, (b) after start of tumor regrowth, (c) immunization with long-life memory cells, (d) induction of diminished tumor growth rate. dashed = killer cells, solid = tumor cells.